

DOI: https://dx.doi.org/10.51227/ojafr.2023.8

EFFECTS OF DIFFERENT PROCESSING METHODS ON NUTRIENT CONTENTS AND ACCEPTABILITY OF HOG PLUM (Spondias mombin Linn.) LEAF BY WEST AFRICAN DWARF SHEEP

Olusegun Oyebade IKUSIKA¹, Adejoke Adeneye MAKO², and Thando MPENDULO¹

¹Department of Livestock and Pasture Sciences, Faculty of Science and Agriculture, University of Fort Hare, Alice, South Africa ²Department of Agricultural Science Education, Tai Solarin University of Education, Ijagun PMB 2118, Ijebu-Ode, Ogun State. Nigeria

ABSTRACT: Three experiments were conducted to evaluate the effect of processing method on leaves from Spondias mombin tree as fodder for ruminants in the tropics. The leaves were subjected to three different physical processing methods; T1 control (fresh but air drying), T2 (fresh but soaked in ordinary water for 24 h then air drying), and T3 (fresh but soaked in water at 50°C for 20 min then air drying). Nutrient and secondary metabolites content were determined in experiment 1. In experiment II, the Coefficient of preference (CoP) was determined. In vitro gas production was used to predict metabolizable energy (ME), organic matter digestibility (OMD), short-chain fatty acid (SCFA) and methane (CH4) of S. mombin leave with different processing methods in experiment 111. Results revealed significant differences in the chemical composition of S. mombin leaf subjected to different processing methods. The dry matter value was highest in S. mombin leaves soaked in hot water (90.22%), and lowest in S. mombin leaves soaked in water at room temperature (85.05%). Crude protein was highest in leaf processed with hot water (11.25 %) and lowest in control (9.59 %). No significant variations were observed for minerals and anti-nutrients investigated. The Vitamin content of leaves of S. mombin tree with various processing methods differed significantly except for vitamin E. The preference coefficient value was greater in leaves soaked at 50°C for 20 mins than leaves from the other processing method and control. All leaves of S. mombin tree from all processing methods considered in this study were acceptable to the animals, but leaves soaked in 50°C for 20 mins were most preferred. The in vitro gas production parameters and characteristics were not significantly different. In conclusion, S. mombin leaf subjected to 50 °C for 20 mins is more advantageous as forage in animal nutrition than unprocessed.

RESEARCH ARTICLE
PII: S222877012200008-12
Received: June 14, 2022
Revised: January 5, 2023
Accepted: January 17, 2023

Keywords: In vitro gas production, Nutritional value, Processing method; Ruminant, Spondias mombin leaf.

INTRODUCTION

The potential of browsing plants as an alternative fodder resource in ruminant nutrition has attracted the attention of researchers worldwide (Mbatha and Bakare, 2018; Medjekal et al., 2020). Several indigenous and exotic browse species have been investigated and evaluated for inclusion in ruminant feeding systems. Unfortunately, farmers' adoption of most of these species faced several challenges, such as pests and disease attacks and the presence of anti-nutritional factors. Therefore, there is the need for continuous screening of browse plants to identify those with good potential as supplements for livestock fodder and which could serve as alternatives to those species that have already been evaluated (Fadiyim et al., 2011). Spondias mombin, popularly known as the Caja plant, belongs to the family Anacardiaceae, native to the tropical Americas and have naturalized in many parts of Africa and Southeast Asia (Duarte and Paull, 2015). It is predominant in all Brazilian regions and West Africa (Sofowora, 2013). Spondias mombin, a rainforest plant, is commonly found in Nigeria's southwest and coastal region (Fadimu et al., 2018). The leaves are used for animal feed during the drought. Various parts of the plant are being used in traditional medicine (Tabuti et al., 2010).

Similarly, Cordeiro et al. (2021) reported that the leaves of *S. mombin* have antimicrobial and antioxidant substances such as tannins, saponins, resins, sterols, triterpenes, flavonoids, and alkaloids. Njoku and Akumefula (2007) report that the leaves of *S. mombin* are potential feedstuff for ruminants because of their richness in crude protein, vitamins, and minerals. Igwe et al. (2010) reported that the crude protein and fibre content from three different species of Spondias mombin ranged from 11.04 to 14.23% and 8.93 to 10.51%, respectively. This crude protein (CP) content compares well with many other browsing plants. Though, biologically active constituent in plants with potential as animal feeding stuff has been a concern. The treatments to which these plants are subjected could have a significant effect on the availability or unavailability of these constituents to animals (Schmitt et al., 2020; Ntalo et al., 2022).

The use of physical treatment of leaves and seeds in livestock feeds to reduce secondary metabolites has been explored by many researchers. Osum et al. (2013) reported a significant (P < 0.05) decrease of micronutrients in the extracts, while it caused a significant (P < 0.05) increase in the protein, fat, and ash content of the oven-dried leaves of *Vitex doniana* subjected to different processing methods. Sallau et al. (2012) also observed an 88.10%, 80.95% and

Email: Oikusika@ufh.ac.za

Supporting Information

61.90% reduction of cyanide in the leaf of *Moringa oleifera* after subjection to boiling, simmering, and blanching, respectively. Similarly, following boiling, the trypsin inhibitor in *Arachis hypogaea* L. (groundnut) lowered to 0.09TUI/g from 0.12TUI/g, while the oxalate content also reduced from 3.04 mg/g to 2.62 mg/g. When Asparagus beans (*Vigna Sesquipedis*) were soaked in water, the alkaloids reduced from 0.34 to 0.28%, phytate from 0.18 to 0.09, tannin from 0.23 to 0.09%, trypsin inhibitor from 13.82 to 9.41 TUI/100g: HCN from 8.63 to 5.68% and Saponin from 0.42 to 0.24% (Nwosu, 2010). Most previous studies explored the medicinal benefits of this plant; however, to the best of our knowledge, there are no studies that consider the effect of processing method on the nutritional value and preferences of *S. mombin* leaf for ruminant nutrition.

Therefore, considering this plant's potential to be a browsing plant, this study is designed to evaluate the effect of processing methods on the nutritional value and acceptability of *Spondias mombin* leaves by West Africa Dwarf sheep.

MATERIAL AND METHODS

Ethical approval: Institutional Animal Care and Use Committee Statement (I.A.C.U.C.)

Routine care and experimental protocols used in this study were approved by the Animal Science Unit in Department of Agricultural Science, of Tai Solarin University of Education, Ijagun. Ijebu-Ode, Nigeria, and conformed to published guidelines for ethical conduct and reporting of research in animal science (Jarvis et al., 2005; Kilkenny et al., 2010).

Harvesting of leaves and identification

Spondias mombin leaves were harvested from Spondias mombin trees located at the botanical garden of Tai Solarin University. Ijebu Ode, Nigeria and authenticated at the Herbarium unit of the Department of Agriculture at the same University.

Processing techniques

About 15 kg of the leaves were harvested, washed, and processed using different methods for nutrient content evaluation, acceptability, and in vitro gas production. The following processing techniques were used. Fresh and air drying for 48 h (control, T1), Fresh but soaking in water for 24 hours, then air-drying for 48 h (T2), and Fresh but soaking in a water bath of 50 °C for 20 mins, then air drying for 2-days (T3) as described in the review of Samtiya et al. (2020). Each processing method processed about 5kg of *Spondias mombin* leaves for the nutritional content, acceptability, and in vitro gas production experiments. After this, the leaf samples from all treatments were packed into sealed nylon, labelled accordingly, and kept for experimental use.

Chemical analysis

Dry matter, crude protein, crude fibre, ether extract and total ash of samples were analyzed in triplicates using the standard procedure described in AOAC (2012). The neutral detergent fibre (NDF), acid detergent fibre (ADF) and acid detergent lignin (ADL) were determined by the Van Soest (1995) method.

Analysis of minerals

A total of ten minerals were analyzed. All samples were digested with HN03 / HClO3 mixtures (nitric acid and perchloric acid) (20:5 v/v). The digest was 100 ml in a standard volumetric flask with deionized water. Ca, Na, K, Fe, Cu, Mn, Zn, Mg and Pb in the digest were determined with the atomic absorption spectrophotometer model 420 (Gallenkemp and Co. Ltd). Phosphorus in the digest was estimated with vanadomolybdate solution. The colour that developed was read with a spectrophotometer at 420 nm.

Quantitative determination of Tannin, Saponin, Oxalate and Phytate

Tannin contents were determined as described by Bohm and Kocipai-Abyazan (1994). Peng and Kobayasli (1995) method was used for saponin analysis. While Oxalate and Phytate contents were determined as described by Oke (1969) and Maga (1983).

Determination of Vitamin content

Vitamins A and E were determined as described by AOAC (2000). The water-soluble Vitamin B was determined as follows: B1 as described by Poornima and Ravishankar (2009), B2 as described by Uraku et al. (2014) and B3 as described by Scalar (2000). Vitamin C was determined as described by Hussain et al. (2006).

Acceptability study

Sixteen West African Dwarf sheep of 18±0.5 kg average body weight was individually housed in pen, 1.5×1.5 m in size, using a complete randomized design with four animals per treatment. Each pen has 3 different feeding troughs (150 × 60 cm each) with one water trough. Leaves (about 1 kg) from each processing method were placed in a separate feeding trough and were strategically placed in the pen in the form of cafeteria feeding, with animal having free access to each of the three feeding troughs (Mako, 2009). The feed preference study lasted for 10 days, excluding a week for the

animals' adaptation to the leaves from different processing methods. The location of feeders was changed every day to prevent the adaptation of the animal to a particular feeding trough. The feeding was allowed from 08:00 to 16:00 hr daily.

The feed consumed was determined by deducting the feed refusal from the quantity offered. Dried samples (about 200 g) of each processing method taken during the 10-day acceptability trial were used to determine the dry matter content. The leaves of processing method preferred were assessed from the Coefficient of preference (CoP) value calculated from the ratio between the intakes of each processing method divided by the average intake of all processing methods (Mako and Babayemi, 2008). Thus, the plant was acceptable, provided the CoP was greater than 1.

In vitro gas production

This was carried out using the method described by Menke and Steingass (1988). A 200 mg milled leaf sample of each processing method was incubated in triplicate with buffered rumen fluid in 120 ml calibrated glass syringes. A total of 12 ewes of West Africa Dwarf sheep were fed concentrate consisting of 40% corn bran, 35% wheat offal, 20% palm kernel cake, 4% oyster shell, 0.5% salt and 0.5% growers' premix for seven days prior to the collection of rumen liquor. Before morning feeding, rumen fluid was collected from the 12 ewes using a suction tube into a pre-heated (39 °C) thermos flask. This was mixed with the buffered mineral solution in the ratio of 1:2 under continuous stirring and flushing with carbon dioxide. Then 30 ml inoculums containing cheesecloth strained rumen liquor and buffer (NaHC03 + Na2HP04 + KCl + NaCl + MgS04. 7H20 + CaCl2. 2H20) were dispensed into a pre-heated sample containing blank syringes and incubated in a water bath maintained at 39 °C. The reading of gas volume was recorded after 3, 6, 9,12,15,18 and 24 h of incubation. The average volume of gas produced from the blanks was deducted from the gas produced per sample. The volume of gas produced at intervals was plotted against the incubation time. From the Graph 1, the gas production characteristics were calculated using the equation, Y= a+b (1-e-ct) described by Orskov and McDonald (1979), where Y= volume of gas produced at the time (t), a = intercept (gas produced from insoluble fraction), c = gas production rate constant for the insoluble fraction (b), and t = incubation time.

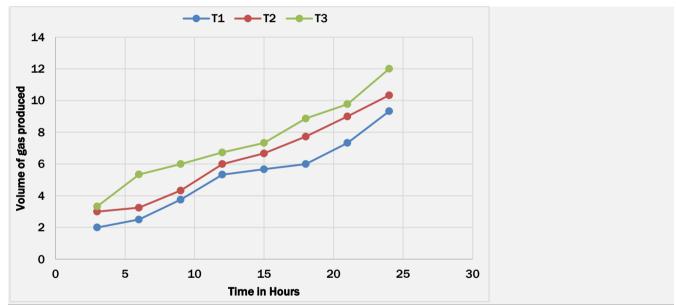


Figure 1 - Gas Production (ml/200mg D.M.) of *Spondias mombin* leaves subjected to different physical processing methods.

The methane gas component was measured at the end of the fermentation period by injecting 4.0 ml of NaOH (10 M) into each syringe containing the incubated samples, following the technique described by Fievez et al. (2005). The measured methane gas volume was related to its respective total gas volume to estimate the methanogenic potential of the digestible OM (Moss et al., 2000). The volume of gas produced after 15 h of incubation was used as an index of energy content and organic matter digestibility (0.M.D.) as described by Menke and Steingass (1988) and short-chain fatty acid (S.C.F.A.) production according to Negesse et al. (2009).

The metabolizable energy (M.E., M.J./Kg D.M.) and organic matter digestibility (O.M.D., %) were estimated as established by Menke and Steingass (1988), and short-chain fatty acids (SCFA, mmol) was calculated as reported (Getachew et al., 1999).

ME = 2.20 + 0.136*GV + 0.057*CP + 0.0029*CF

OMD = 14.88 + 0.889GV + 0.45CP + 0.651XA

SCFA = 0.0239*GV - 0.0601

Where GV, CP, C.F. and X.A. are net gas productions (ml /200 mg D.M.), crude protein, crude fibre, and ash of the incubated samples.

Statistical analysis

Data obtained were analyzed using SAS's general linear model (GLM) procedure- 2012. Single-factor analysis of variance (ANOVA) was used to assess the effects of the processing method of *Spondias mombin* leaf on nutrient composition, coefficient of preference, in vitro gas production, O.M.D., ME and S.C.F.A. with the following model: Yij = μ + Bi + Eij. Where: Yij is an observation, μ is the overall mean, Bi is the effect of leaf growth stages, and Eij is the experimental error. Significant means were separated by Duncan's (1955) Multiple Range Test. Differences between means were considered statistically significant, P<0.05.

RESULTS

The result of the chemical composition is presented in Table 1. It was observed that all parameters examined varied significantly (P<0.05). The dry matter ranged from 85.05 – 90.22 %, with leaves soaked in water at room temperature for 24 hours having the lowest while leaves soaked in 50 °C for 20 mins the highest value. The crude protein content ranged from 9.59-to 11.25%. The macro and micro mineral content of S. *mombin* leaves subjected to different physical processing methods are presented in Table 2. The result revealed that all leaves from different processing methods contained all minerals investigated. There were no significant variations among the leaves from different processing methods for all the minerals investigated.

Presented in Table 3 is the anti-nutritional content of *Spondias mombin* leaves subjected to different physical processing methods. All the physical processing methods were implicated for all the antinutrients investigated. No significant variation occurred among the different processing methods. The tannin, oxalate, saponin and phytate levels ranged from 0.27-0.88; 0.392-0.432; 0.281-0.397 and 0.6-0.623 %, respectively.

Table 4 presents the vitamin content of leaves subjected to different physical processing methods. All vitamins investigated differed significantly (P<0.05) except vitamin E. In all the vitamins analyzed, the S. mombin leaves subjected to hot water (50 °C) for 20 mins had the lowest values while the control had the highest values. Table 5 presents the Coefficient of preference (CoP) of Spondias mombin leaves exposed to different physical processing methods by sheep. Table 6 shows the *in vitro* gas production parameters of Spondias mombin leaves exposed to different physical processing methods. No significant differences were observed for metabolizable energy (ME), organic matter digestibility (OMD), short-chain fatty acid (SCFA) and methane (CH₄). They ranged from 3.89-4.52 MJ/Kg D.M., 37.13-39.01%, 0.15-0.22 µmol, and 3.50-6.00 ml soaked in hot water for 20 mins, soaked in water at room temperature and the control, respectively.

Figure 1 presents the gas production pattern of the different processing methods of Spondias mombin leaves incubated for 24 hr.

Physical processing methods	T1	T2	тз	S.E.M.
Parameters	••	12	15	J.L.IVI.
Dry matter (DM)	85.05c	89.34b	90.22a	0.42
Crude protein (CP)	9.59∘	10.28b	11.25a	0.33
Crude fibre	17.18a	16.64 ^b	15.23°	0.20
Ether extract	8.24a	7.95⁵	7.50∘	0.30
Ash	10.00a	9.79⁵	8.25c	0.20
Neutral detergent fibre	46.48c	57.94b	55.01a	0.32
Acid detergent fibre	33.63°	35.52b	45.67ª	0.40
Acid detergent lignin	10.98a	10.24b	9.56c	0.30

Table 2 - Macro and micro r methods	nineral co	ntent of	Spondias	mombir	leaves	subjected	l to diffe	erent phy	sical pro	cessing
Processing methods		Mac	rominerals	(%)			Micro n	ninerals (r	ng/kg)	
	Ca	Р	K	Na	Mg	Fe	Zn	Cu	Mn	Pb
T1	1.230	0.22	2.156	1.21	0.41	569.0	58.5	13.3	22.4	11.2
T2	1.330	0.17	1.70	1.17	0.38	551.5	54.3	11.0	20.4	9.1
Т3	1.341	0. 13	1.65	1.06	0.26	546.1	53.3	10.8	17.6	8.4
SEM	0.29	0.15	0.13	0.30	0.29	10.9	2.5	0.8	7.3	0.9
SEM= Standard Error of the mean (*P<0.05)									

Table 3 - Secondary Metabolites (%) content of Spondias mombin leaf subjected to different physical processing methods Physical processing methods **T1** T2 **T3** S.E.M. **Parameters** Tannin 0.88 0.44 0.27 0.41 **Oxalate** 0.432 0.408 0.392 0.42 Saponin 0.397 0.368 0.281 0.42 **Phytate** 0.623 0.600 0.42 0.617 SEM= Standard Error of the mean (*P<0.05)

Table 4 - Vitamin content (mg/100 g) of Spondias mombin subjected to different physical processing methods							
Physical processing method Vitamins	ts T1	T2	Т3	S.E.M.			
A	5.72a	5.10b	4.38c	0.01			
B1	0.060a	0.040b	0.021c	0.001			
B2	0.31a	0.281 ^b	0.223c	0.001			
В3	3.87a	3.54b	3.21c	0.02			
C	19.01 ^a	17.21 ^b	14.43°	0.21			
E	1.12 ^a	1 .0 1 ^a	1.10a	0.001			

Tak	le 5 - Mean of daily intake of leaves from	different parts of	Spondias mombin	and the Coefficient	of preference by
WA	D sheep.				

Processing methods	Mean dally (kg/D.M.) consumption by all animals	Coefficient of preference	Ranking
T1	3.26	1.85	3
T2	3.40	1.89	2
Т3	4.23	2.05	1

				ssing methods
In vitro parameters	ME	SCFA	OMD	CH₄
Methods	(MJ/Kg DM)	(µmol)	(%)	(ml/200mg DM)
T1	3.89	0.15	37.13	6.00
T2	4.12	0.18	38.78	4.15
Т3	4.52	0.22	39.01	3.50
SEM	0.24	0.04	1.75	2.00

Table 7 - In vitro characteristics of Spondias mombin leaves subjected to different physical processing methods.
--

In vitro characteristics	а	b	С	a+b
Methods	(ml/200 mg DM)	(ml/200 mg DM)	(ml/h)	(ml/200 mg DM)
T1	1.10	14.45	0.10	16.01
T2	1.30	15.13	0.10	17.23
Т3	1.43	18.06	0.11	19.13
SEM	0.1	1.05	0.02	1.52

a= soluble degradable fraction; b= insoluble degradable fraction; a+b= Potential degradability; c= rate of degradation; SEM= standard error of mean (*P<0.05)

DISCUSSION

The proximate analysis results quickly estimate the nutrient content of leaves or feedstuff and show the potential and clues for further research. Though, the proximate analysis of the feed may not be the true reflection of the nutritional value of such feedstuff. It provides the primary guideline in screening feedstuff with the potential of being a browse plant for ruminant animals.

The dry matter (DM) and the crude protein values obtained in the study are more in the processed leaves than in control. This result agrees with Adebayo et al. (2014) but is lower than the 15.44% CP reported by Omoniyi et al. (2013). The same trend as the dry matter and crude protein was observed for ADF and NDF. These values are lower and at variance with the values reported by Igwe et al. (2010) but in agreement with the values reported by Adebayo et al. (2014). The CP content is more than that proposed as the minimum requirement for growth (113g C.P./Kg D.M.) and lactation (120 CP/Kg D.M.) in ruminants (NRC, 2001). This makes Spondias mombin leaves a good source of protein supplement for low-quality roughage. The values of NDF for physically processed leaves are within the recommended limit of 60.00% guaranteed for forage intake by ruminants (Wanapat et al., 2013). The ash content of the processed leaves was lower than the control but was still high despite the treatment effect. This suggests that many inorganic elements are contained in the leaves (Akinmoladun, 2018; Mishra et al., 2022). The macro and micro mineral content of S.mombin leaves subjected to different physical processing methods revealed that all leaves from different processing methods contained all minerals investigated. There were no significant variations among the leaf from different processing methods for all the minerals investigated. However, all the minerals investigated have higher values in control than the processed except for Calcium. This implies that most of the minerals analyzed in this study are vulnerable to water treatment, reducing them when subjected to water treatment. Yet, the values obtained in the processed leaves are within the recommended limit for the proper functioning of the body system and agree with the values reported for Spondias mombin by Ayoola et al. (2010).

Tannins are plant polyphenols that can form complexes with metal ions and macromolecules such as proteins and polysaccharides, thereby protecting them from ruminal degradation (Akubugwo and Ugbogu, 2007; Siemińska-Kuczer et al., 2022). Mlambo et al. (2015) report that 2-3% of tannin in ruminants' diets is beneficial because it helps in reducing rumen protein degradation by the protein-tannin complex formed. They can reduce feed intake, feed efficiency and weight gain when the value is above 60 to 100g/kg (Ogbe and George, 2012). Saponin suppresses methanogenesis, a significant energy loss to animals (Wang et al., 2009). Agbaire (2012) observed that Saponin higher than 10% in ruminant diets could result in gastroenteritis, diarrhoea, and dysentery. Oxalate can form complexes with most essential trace elements, making them unavailable for enzymatic activities and other metabolic processes. High doses of oxalic acid cause corrosive gastroenteritis, shock, convulsive symptoms, and renal damage (Eneobong, 2001). Phytic acid inhibits the absorption and utilization of some mineral elements (Eneobong, 2001). In this study, the values of all phytochemicals investigated are lower in the processed treatments than in the control. This implies that subjecting S. mombin leaves to water for 24 at room temperature or to hot water (50 °C) for 20 mins would significantly reduce the secondary metabolites considered in this study. Therefore, S. mombin leaves treated by physical methods in this study lower the antinutrients appreciably without adversely affecting the proximate composition. All phytochemicals investigated were within the safe limit for ruminant consumption, as reported by Teferedgne (2000). This makes the values of these phytochemicals in all the different processing methods in this present study beneficial. Vitamins are organic molecules that are essential micronutrients which an organism needs in small quantities for an efficient functioning of its metabolism system. Vitamins quantity produced via the body is insufficient. Therefore, sources through diet are essential. The values of all the investigated vitamins are lower in the processed leaves compared with the control. This could be because most of these vitamins, except vitamin E, are water-soluble vitamins. However, the values fall within the recommended levels for ruminant animals, as Maduka et al. (2014) reported. The high values of vitamins observed showed that the plant has a high nutritive value which could attenuate physiological oxidative stress due to its high concentration of vitamin C and E (Maduka et al., 2014). Also, feeding this plant to ruminants will boost such animals' immune and reproductive systems because of the high-level vitamin C and E content.

The nutritional content of a feed is crucial; however, an animal's acceptance of such feed is more vital. Hence, this aspect of the study was looked into. The animals' co-efficient of preference (CoP) showed that all leaves subjected to different physical processing methods were acceptable to the animals because the CoP was greater than unity (Mako and Babayemi, 2008). Many factors may influence the acceptability of feed by ruminants. Provenza and Cinocotta (1994) reported that physical plant structure and chemical composition are the most vital factors influencing animals' preference for feed. The crude protein of the leaf subjected to soaking in hot water for 20 mins was the highest. This might be the reason why it is most preferred. Also, this could be because this treatment contains the lowest values of phytochemicals like tannins that can affect the palatability of browse plants compared to others. Similarly, the action of a short time soaking in hot water could make the leaves succulent and soft; hence the most preferred compared with others.

In combination with in vitro digestibility and ME content, chemical composition can be considered valuable indicators for preliminary evaluation of the potential nutritive value of forages (Kafilzadeh and Heidary, 2013). The values of ME, OMD, and SCFA obtained here indicate that animals will obtain energy from the S. mombin leaves subjected to different physical processing methods. Methane production is an energy loss to ruminants. Also, it has environmental implications as a significant contributor to greenhouse gas emissions (USEPA, 2014). Feedstuff that shows a high capacity for gas production is also synonymous with high methane production (Njidda, 2010), hence the reason for high methane production in control compared with others. The result obtained in this present study aligned with the 4.74 MJ/Kg D.M.; 38.03 % and 0.22 µmol reported for Spondias mombin leaf by Omoniyi et al. (2013), but lower and at variance with the report for Spondias mombin leaf by Ogunbosoye and Babayemi (2010). This variance could be due to different processing methods used.

The same trend as in vitro gas production parameters was observed for a, b, a+b and c. These results are lower and at variance than Ogunbosoye and Babayemi (2010) report for Spondias mombin leaf.

The gas production pattern reflects the effectiveness and extent of forages' degradability because forages with high ruminal degradability of dry matter tend to have high gas production (Njidda and Nasiru, 2010). A high lignin level negatively influences the degradation of fibre and non-fibre (Mizubiti et al., 2011). The lignin content obtained in the control treatment in this study is high, hence the low volume of gas produced. There was a steady increase in the gas production for 24 hours; no significant variations were observed in the gas volume produced at every interval. Many factors may determine the amount of gas produced during fermentation, depending on the nature and level of fibre, the presence of secondary metabolites and the potency of rumen liquor (Babayemi et al., 2004). The values observed in this present study agree with the report for Spondias mombin leaf by Omoniyi et al. (2013) but are lower and at variance than the 43 – 175 ml reported by Ogunbosoye and Babayemi (2010). This could be attributed to the processing methods implored in the research. The processed treatments have lower lignin content compared with unprocessed. The low volume of gas production of the Spondias mombin leaf in control connotes the low digestibility of the leaf, which may be due to the higher values of anti-nutritional factors and fibre compared with the processed.

CONCLUSION

The nutrient, antinutrient, mineral, vitamin content, and *in vitro* gas production fermentation revealed that Hog plum (*Spondias mombin*) leaf has potential forage for ruminants when subjected to physical processing methods such as soaking in hot water (50 °C) for 20 mins. In addition, animals prefer Hog plum leaves soaked in hot water (50 °C) for 20 min. compared with ordinary air-dried leaves before feeding.

DECLARATIONS

Corresponding author

Olusegun Oyebade IKUSIKA; Email: Oikusika@ufh.ac.za

Authors' contribution

Conceptualization, investigation, writing, data collection and methodology were carried out by Ikusika Olusegun and Adejoke Mako. While Thando Mpendulo and Ikusika did the statistical analysis, supervision, and validation of the research work.

Acknowledgement

The authors wish to acknowledge the Department of Livestock and Pasture Science, University of Fort, South Africa, Govan Mbeki Research and Development Centre, University of Fort Hare and Tai Solarin University of Education, Ijebu-Ode, Ogun State, Nigeria for providing the platform and enabling environment for this research.

Funding

This research received no external funding.

Consent to publish

Not applicable.

Disclosure of statement

The authors report no conflict of interest.

Data availability statement

Data Will be available on request.

REFERENCES

Adebayo BJ, Omoniyi LA, Isah OA, Olorunsola RA, Akinfemi A, Olayemi WA, et al. (2014). Chemical composition of Pennisetum purpureum, Spondias mombin and Ageratum conyzoides used as feed for goats and sheep at Odoragunshin of Epe Local Government, Lagos State. Proceedings of 39th Annual Conference of the Nigerian Society for Animal Production (N.S.A.P.), Nigeria, pp. 181-184.

Akinmoladun OF, Sabi RS, and Adedayo OT (2018). Toxicological evaluation of graded levels of freshly harvested bamboo (*Bambusa arundinacea*) and Tridax (*Tridax procumbens*) leaves on blood chemistry of rabbits. Nigerian Journal of Animal Production, 45(1): 183-197. DOI: https://doi.org/10.51791/njap.v45i1.331

Akubugwo IE and Ugbogu AE (2007). Physiochemical studies on oils from five selected Nigerian plant seeds. Pakistan Journal of Nutrition. 6 (6): 75-78. DOI: https://dx.doi.org/10.3923/pjn.2007.75.78

Agbaire PO (2012). Levels of anti-nutritional factors in some common leafy edible vegetables of Southern Nigeria. African Journal of Food Science and Technology, 3(4): 99–101. Available online https://www.interesjournals.org/articles/levels-of-antinutritional-factors-in-some-commonleafy-edible-vegetables-of-southern-nigeria.pdf

Ahamefule FO, Obua BE, Ibeawuchi JA and Udosen NR (2006). The Nutritive Value of Some Plants Browsed by Cattle in Umudike, Southeastern Nigeria. Pakistan Journal of Nutrition, 5 (5): 404-409. DOI: https://dx.doi.org/10.3923/pjn.2006.404.409v

AOAC (2000). Official Method of Analysis Association of Analytical Chemist International. 17th Edition, Horowitz, Maryland.

- AOAC (2012). Official Methods of Analysis, 19th ed. (Association of Officials Analytical Chemists, Arlington, VA).
- Ayoola PB, Adeyeye A and Onawunmi OO (2010). Trace elements and major mineral evaluation of Spondias mombin, Vernonia amygdalina and Momordica charantia leave. Pakistan Journal of Nutrition, 9 (8): 755-758. DOI: https://dx.doi.org/10.3923/pjn.2010.755.758
- Babayemi OJ, Demeyer KD and Fievez V (2004). Nutritional value and qualitative assessment of secondary compounds in seeds of eight tropical browse, shrub and pulse legumes. Communications in Agricultural and Applied Biological Sciences, 69(1): 103–110. https://europepmc.org/article/med/15560266
- Bohm BA and Kocipai AC (1994). Flavonoid and Condensed tannins from leaves of Vaccinum raticulation and Vaccinum calcyimium. Pacific Science Journal, 48: 458-463, Google Scholar
- Cordeiro TO, Mendonça AKP, Gomes MS, Ferreira de Lima EL, Macedo Costa MR, Costa de Lima K, Alves Uchôa Lins RD (2021). Antimicrobial action of the hydroethanolic extract of Spondias mombin L. against oral bacteria of the genus Streptococcus. Research, Society and Development, 10(1): e39310111539. https://rsdjournal.org/index.php/rsd/article/view/11539
- Duarte O, and Paull R (2015). Exotic fruits and nuts of the new world. Cabi Publication, UK. Pp. 332. https://www.cabdirect.org/cabdirect/abstract/20153017861; https://dx.doi.org/10.1079/9781780645056.0000
- Eneobong HN (2001). Eating Right (A Nutrition Guide). Zoometer print communications Ltd. Nigeria.
- Fadimu GL, Sanni LS, Adebowale AR, Kareem S, Sobukola OP, Kajihausa O, Saghir A, Siwoku B, Akinsanya A, Adenekan MK (2018). Effect of drying methods on the chemical composition, colour, functional and pasting properties of plantain (*Musa parasidiaca*) flour. Croatian Journal of Food Technology, Biotechnology and Nutrition, 13 (1-2): 38-43. DOI: https://doi.org/10.31895/hcptbn.13.1-2.2
- Fadiyim AA, Fajemisin AN and Alokan JA (2011). Chemical composition of selected browse plants and their acceptability by West African Dwarf sheep. Livestock Research for Rural Development, 23: Article #256. http://www.lrrd.cipav.org.co/lrrd23/12/fadi23256.htm
- Getachew G, Makkar HPS and Becker K (1999). Stoichiometric relationship between short-chain fatty and In vitro gas production in presence and absence of polyethene glycol for tannin-containing browses. E.A.A.P. satellite symposium gas production; Fermentation kinetics for feed evaluation and to assess microbial activity. August 18- 19. Wageningen, Netherlands.
- Hussain I, Saleem M, Igbal Y and Khalil SJ (2006). Comparison of Vitamin C contents in Commercial Tea Brands and Fresh Tea leaves. Journal of the Chemical Society of Pakistan, 28: 421-425. https://jcsp.org.pk/issueDetail.aspx?aid=1fc23f6c-44af-4a57-92c0-61f696b93efc
- Igwe CU, Onyeze GOC, Onwuliri VA, Osuagwu CG and Ojiako AO (2010). Evaluation of the chemical composition of the leaf of Spondias mombin
 Linn from Nigeria. Australian Journal of Basic and Applied Sciences, 4 (5): 706-710.

 https://www.cabdirect.org/cabdirect/abstract/20103359541
- Jarvis SC, Day JEL and Reed B (2005). Ethical guidelines for research in animal science. Published by The Norwegian National Research Ethics Committee on 8/7/2019
- Kafilzadeh F and Heidary N (2013). Chemical composition, *in vitro* digestibility and kinetics of fermentation of whole-crop forage from 18 different varieties of oat (*Avena sativa* L.). Journal of Applied Animal Research, 41 (1): 61-68. DOI: DOI: https://doi.org/10.1080/09712119.2012.739084
- Kilkenny C, Browne WJ, Cuthill IC, Emerson M and Altma DG (2010). Improving Bioscience Research Reporting: The ARRIVE Guidelines for Reporting Animal. PLOS Biology, 8(6): e1000412. DOI: https://doi.org/10.1371/journal.pbio.1000412
- Maduka HCC, Okpogba AN, Ugwu CE, Dike CC, Ogueche PN, Onwuzurike DT and et al. (2014). Phytochemical, antioxidant and microbial inhibitory effect of Spondias mombin leaf, stem and bark extracts. IOSR Journal of Pharmacy and Biological Sciences, 9(2): 14-17. Article-link; Google Scholar
- Maga JA (1983). Phytate: its chemistry: occurrence, food interactions, nutritional significance and method of analysis. Journal of Agricultural Food Chemistry, 30(1): 1-9. DOI: https://doi.org/10.1021/jf00109a001
- Mako AA and Babayemi OJ (2008). Nutritional evaluation of water hyacinth as ruminant feed to curtail its environmental menace in Nigeria. African Journal of Livestock Extension, 6: 48–53. https://www.ajol.info/index.php/ajlex/article/view/50092
- Mako AA (2009). Evaluation of water hyacinth (*Eichhornia crassipes Mart*.Solms Laubach) as a potential feed for the West African Dwarf goats. PhD. Thesis. Department of Animal Science. The University of Ibadan.
- Mbatha KR, and Bakare AG (2018). Browse silage as potential feed for captive wild ungulates in southern Africa: A review. Animal Nutrition, 4(1): 1-10. DOI: https://doi.org/10.1016/j.aninu.2017.12.003
- Medjekal S, Ghadbane M, Boufennara S, Bebderradji L, Bodas R, Bousseboua H, et al. (2020). Chemical composition, in situ degradation, and fermentation kinetics of some browse plant species collected from Algerian arid and semi-arid areas. Journal of Rangeland Science, 10(2): 188-203. https://rangeland.borujerd.iau.ir/article_669014_35aab5483de7e3176fd79c4245df93fd.pdf
- Menke KH and Steingass H (1988). Estimation of the energetic feed value from chemical analysis and in vitro gas production using rumen fluid. Animal Research Development, 28: 7-55. DOI: https://agris.fao.org/agris-search/search.do?recordID=DE88A022488
- Mishra P, Kumar R, and Rai AK (2022). Analysis of tendu (Diospyros melanoxylon) leaf using spectroscopic techniques. National Academy Science Letters, 45(1):91-94. DOI: https://doi.org/10.1007/s40009-021-01075-6
- Mizubiti IY, Ribeiro EA, Pereira ES, Pinto AP, Franco ALC, Syperreck MA and Muniz EB (2011). In vitro rumen fermentation kinetics of some coproducts generated by gas production technology in the bio-diesel production chain. Semina: Ciencais Agrairias, 32(Supplemento): 2021–2028. https://agris.fao.org/agris-search/search.do?recordID=DJ2012067959
- Mlambo V, Marume U and Gajana CS (2015). Utility of the browser's behavioural and physiological strategies in coping with dietary tannins: Are exogenous tannin-inactivating treatments necessary? South African Journal of Animal Science, 45(5): 441-451. DOI: http://dx.doi.org/10.4314/sajas.v45i5.1
- Mtengeti EJ and Mhelela A (2006). Screening of indigenous browse species in semi-arid central Tanzania. A case of Gairo division. Livestock Research for Rural Development 18(8):108. http://www.lrrd.org/lrrd18/8/mten18108.htm
- Negesse T, Makkar HP, and Becker K (2009). Nutritive value of some non-conventional feed resources of Ethiopia determined by chemical analyses and an in vitro gas method. Animal Feed Science and Technology, 154(3-4):204-217. DOI: https://doi.org/10.1016/j.anifeedsci.2009.09.010
- Njidda AA (2010). Determining Dry matter degradability of some Semi-Arid Browse species of North-Eastern Nigeria using in vitro Technique. Nigerian Journal of Basic and Applied Science 18 (2): 160 167. Available online at http://ajol.info/index.php/njbas/index
- Njidda AA and Nasiru A (2010). In vitro gas production and dry matter digestibility of tannin-containing forages of the Semi-arid region of Northern Eastern Nigeria. Pakistan Journal of Nutrition, 9 (1): 60 66. DOI: https://dx.doi.org/10.3923/pjn.2010.60.66
- NRC (2001). Nutrient requirements for dairy cattle seventh revised edition. National Research Council. National Academy of Science Press. Washington DC. U.S.

- Ntalo M, Ravhuhali KE, Moyo B, Hawu O, and Msiza NH (2022). Lantana camara: Poisonous Species and a Potential Browse Species for Goats in Southern Africa—A Review. Sustainability, 14(2):751. DOI: https://doi.org/10.3390/su14020751
- Nwosu JN, Ogueke CC, Owuamanam CI, and Iwouno JO (2010). Functional Properties and Proximate Composition of Asparagus Bean (Vigna Sesquipedalis) as Influenced by Malting. Journal of American Science, 6(9): 376-382. http://www.jofamericanscience.org/journals/amsci/am0609/43 3416am0609 376 382.pdf
- Ogbe AO and George GAC (2012). Nutritional and anti-nutritional content of melon husk: Potential as a feed ingredient in poultry diet. Research Journal of Chemical Sciences, 2(2): 35-39. http://www.isca.in/rjcs/Archives/v2/i2/5.php
- Ogunbosoye DO and Babayemi OJ (2010). Potential values of some non-leguminous browse plants as dry season feed for ruminants in Nigeria. African Journal of Biotechnology 9(18): 2720–2726. https://www.ajol.info/index.php/ajb/article/view/79908
- Oke CL (1969). Oxalic acid in plants and nutrition. World Review of nutritional and Dietetics, New York.
- Omoniyi LA, Isah OA, Taiwo OO, Afolabi AD and Fernadez AJ (2013). Assessment of Nutritive value of some Indigenous Plants Consumed by Ruminants in the Humid and Sub-Humid Region of Nigeria using In vitro Technique. The Pacific Journal of Science and Technology, 14 (1): 413-421. Article link; Google Scholar
- Orskov ER and McDonald I (1979). The estimation of protein degradability in the rumen from incubated measurements weighted according to the rate of passage. Journal of Agricultural Science, 92: 499-503. DOI: https://doi.org/10.1017/S0021859600063048
- Osum F, Okonkwo TM and Okafor GI (2013). Effect of processing methods on the chemical composition of *Vitex doniana* leaf and leaf products. Food Science & Nutrition, 1(3): 241– 245. DOI: https://doi.org/10.1002/fsn3.31
- Peng and Kobayasli (1995). Novel furostanol glycosides from *Allium macrostemon*. Media, 6: 58-61. DOI: https://doi.org/10.1055/s-2006-958000 https://www.thieme-connect.de/products/ejournals/abstract/10.1055/s-2006-958000
- Poornima GN and Ravishankar RV (2009). Evaluation of Phytonutrients and Vitamin Contents in a Wild Yam *Dioscorea belophylla* (prain) Haines. African Journal of Biotechnology, 8: 971-973. https://www.ajol.info/index.php/ajb/article/view/59997
- Provenza FD and Cincotta RP (1994). Forage as a self-organizational learning process; accepting adaptability at the expense of predictability. Hughes, R. N. (ed) Diet selection. Blackwell Scientific publications. Oxford Pp 79 101.
- Uraku AJ, Okala ANC and Ibiam UA (2014). Effect of Spilanthes uliginosa (sw), Ocimum basilicium, Hyptis spicigera and Cymbopogon citrates leaf extracts on Biochemical and Histological Parameters of Mice Exposed to Plasmodium berghei. PhD Thesis submitted to the Department of Biochemistry, Ebonyi State University. Abakaliki. Nigeria.
- Sallau AB, Mada SB, Ibrahim S and Ibrahim U (2012). Effect of boiling, simmering and blanching on the anti-nutritional content of Moringa oleifera leaves. International Journal of Food Nutrition and Safety, 2(1):1-6. https://www.semanticscholar.org/paper/Effect-of-Boiling%2C-Simmering-and-Blanching-on-the-Sallau-Mada/ddc6dc15f318a8e83cd94ac2076ca1728e47eaa4
- Samtiya M, Aluko RE and Dhewa T (2020). Plant food anti-nutritional factors and their reduction strategies: an overview. Food Production, Processing and Nutrition, 2(1): 1-14. DOI: https://doi.org/10.1186/s43014-020-0020-5
- SAS (2012). Statistical Analytical Systems. S.A.S. Version 9.2 user's guide. Carry, NY: SAS Institute.
- Scalar (2000). In: Segregated flow analyzer for Analytical Process Laboratories, Netherland. Pp 45, 55 and 61.
- Sofowora L (2013), Medicinal plants and traditional medicine in Africa, Ibadan; Spectrum Books Ltd.
- Schmitt MH, Shuttleworth A, Shrader AM, Ward D (2020). The role of volatile plant secondary metabolites as pre-ingestive cues and potential toxins dictating diet selection by African elephants. Oikos, 129(1):24-34. DOI: https://doi.org/10.1111/oik.06665
- Siemińska-Kuczer A, Szymańska-Chargot M, and Zdunek A (2022). Recent advances in interactions between polyphenols and plant cell wall polysaccharides as studied using an adsorption technique. Food Chemistry, 373:131487. DOI: https://doi.org/10.1016/j.foodchem.2021.131487
- Tabuti JRS, Kukunda CB, Waako PJ (2010). Medicinal plants used by traditional medicine practitioners in the treatment of tuberculosis and related ailments in Uganda. Journal of ethnobiology and ethnomedicine, 9(1): 1-23. DOI: https://doi.org/10.1186/1746-4269-9-65
- Teferedgne B (2000). New perspectives in the use of tropical plants to improve ruminant nutrition. Proceedings of the Nutrition Society, 59(2): 209-214. DOI: https://doi.org/10.1017/S0029665100000239
- USEPA. (2014). (the United States Environmental Protection Agency). Inventory of US Greenhouse Gas Emissions and Sinks: 1990 2012. Available online: http://www3.epa.gov/climatechange/Downloads/ghgemissions/US-GHG-Inventory-2014-Main-Text.pdf.
- Van Soest PJ (1995). Nutritional Ecology of Ruminants, 2nd edn. Cornell University Press.
- Wanapat M, Pilajun R, Polyorach S, Cherdthrong A, Khejornsart P and Rowlinson P (2013). Effect of Carbohydrate sources and Cottonseed meal level on feed intake, nutrient digestibility, Rumen fermentation and Microbial Protein Synthesis in swamp Buffaloes. Asian-Australas Journal of Animal Science, 26 (7): 52-60. www.ajas.info/journal/view.php?&number=4738
- Wang CJ, Wang SP and Zhou H (2009). Influences of flavomycin, ropadiar and Saponin on nutrient digestibility, rumen fermentation and methane emissions from sheep. Animal Feed Science and Technology, 148: 157-166. https://doi.org/10.1016/j.anifeedsci.2008.03.008